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Relationship between invasion of the *periodontium* by periodontal pathogens and periodontal disease: a systematic review

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ABSTRACT

Background: Bacterial invasion of the periodontal tissues has been suggested as a relevant step in the etiopathogenesis of periodontal disease. However, its exact importance remains to be defined. The present systematic review assessed the scientific evidence concerning the relationship between the quality or quantity of periodontal microbiota in periodontal tissues and development of periodontal disease.

Methods: The databases Medline-PubMed, Cochrane-CENTRAL, ISI Web of Knowledge and SCOPUS were searched, up to January. Studies that reported evaluation of periodontal pathogens invasion on human tissues were selected.

Results: The screening of 440 title/abstracts elected 26 papers for full-text reading. Twenty three papers were subsequently excluded because of insufficient data or a study protocol not related to the objectives of this systematic review. All included studies were case-control studies that evaluated intracellular or adherent bacteria to epithelial cells from periodontal pockets *versus* healthy sulci. Study protocols presented heterogeneity regarding case and control definitions and methodological approaches for microbial identification. No consistent significant differences were found related to the presence/absence or proportion of specific periopathogens across the studies, as only one study found statistically significant differences regarding the presence of *A. actinomycetemcomitans* ($p=0.043$), *T. forsythia* ($p<0.001$), *P. intermedia* ($p<0.001$), *C. ochracea* ($p<0.001$) and *C. rectus* ($p=0.003$) in epithelial cells from periodontal pockets *versus* healthy sulci. All studies reported a larger unspecific bacterial load in or on the epithelial cells taken from a diseased site compared to a healthy sulcus.

Conclusion: The current available data is of low to moderate quality and inconsistent mainly due to study design, poor reporting and methodological diversity. As so, there is insufficient evidence to support or exclude the invasion of periodontal pathogens as a key step in the etiopathogenesis of periodontal disease. Further research is needed.

INTRODUCTION

Over the past 50 years the role of invasion of bacteria in the complex pathogenic process of periodontal disease has undergone cycles of acceptance and rejection. Since Listgarten¹, and its first electron microscopy images of spirochetal infiltration in gingival tissues from patients with necrotizing ulcerative gingivitis, the invasiveness of oral bacteria emerged as a potentially important mechanism that mediates initiation and progression of periodontal disease.

It is not difficult to conjecture that the presence of microorganisms within gingival tissues would not augur good things to the *periodontium*. Tissue invasion allows direct discharge of destructive bacterial products²⁻⁴ and promotes the release of lysosomal contents from neutrophils in the periodontal tissues.^{5,6} Besides that, invasive bacteria seem to have mechanisms to evade host defenses. To be sheltered from the humoral immune surveillance invasive bacteria penetrate and remain within the epithelial cells, in a nutritious environment, where they can replicate and spread to neighboring cells.⁷ Additionally, *in vitro* studies have shown that *Porphyromonas gingivalis* impedes transepithelial migration of neutrophils and prevents epithelial cells from secreting IL-8 in response to bacterial challenge.^{8,9} Several studies have also suggested that periodontal bacteria actively suppress cell-mediated immunity and this, presumably, contributes to periodontal lesion development.¹⁰ Standard periodontal treatment is also undermined as intracellular bacteria are less likely to be physically removed by scaling and root planning¹¹ and are more resistant to antibiotics.¹²

Although there is a theoretical basis for a role of invasion in the etiopathogenesis of periodontal disease and *in vitro* studies that support that bacteria possess the molecular machinery to perform invasion and deceive host defenses⁷, few *in vivo* studies were conducted¹³⁻¹⁶, letting the idea in abeyance throughout the years.

Findings of putative periodontal pathogens in healthy sulci¹⁷, the impossibility to discriminate periodontal disease by microbiologic analysis¹⁸, the inability to categorically refute the possibility that bacteria were not artificially introduced into the tissues during collection or

processing, in some studies¹⁹, and evidence of internalized putative periodontal pathogens in epithelial buccal cells of healthy individuals^{20,21} helped to sustain this doubt.

As the comprehensive knowledge of the whole dynamic of the periodontal interface is crucial for improving diagnostics and setting effective and rational treatments and the exact importance of bacteria invasiveness in the etiopathogenesis of periodontal disease remains unclear there is a need for a systematic assessment of the literature on this topic. The aim of the present systematic review was to assess the existing scientific literature to ascertain the relationship between the quality or quantity of periodontal microbiota in periodontal tissues and periodontal disease.

RESULTS

Search and selection results

The search resulted in 440 unique papers, in which titles and abstracts were screened (for details see Fig. 1). Of these, 26 met the eligibility criteria^{14,22-46} and were selected for full text reading. One study was in Japanese and was excluded.³⁶ Twenty two studies were subsequently excluded for specific reasons: 3 studies^{34,35,37} had a study protocol not related to the objectives of this systematic review, 8 studies^{14,25-27,30,39,40,42} had absence of control group (non-diseased samples), one study³⁸ only presented disease samples from a single patient, 7 studies^{28, 29, 31-33,41,44} hadn't clearly defined inclusion and/or exclusion criteria, and finally three studies^{43, 45, 46} were excluded because they only addressed opportunistic pathogens. Additional hand searching of the reference lists of the selected papers didn't retrieve any additional studies. As such, only three papers were included in the present systematic review.

General trial characteristics and heterogeneity

The three included studies were case-control studies with considerable heterogeneity. In all studies subjects with systemic diseases and antibiotic intake in the previous 6 months were excluded. Two studies (#2, #3) also excluded pregnant women. In two studies (#2, #3) diseased sites were defined as having probing pocket depth (PPD) and clinical attachment lost (CAL) ≥ 4 mm and healthy sites as having PPD ≤ 3 mm and CAL < 4 mm while in the other study (#1) diseased sites were defined as having PPD > 5 , bleeding on probing (BOP) and suppuration and healthy sites as having PPD < 5 with no BOP or suppuration. No information about CAL thresholds was provided in this last study. One study (#2) provided smoking habits information of the participants while in the other two (#1, #3) this information was not reported. Two studies (#1, #3) had separate case and control groups of participants. In one study (#2) each participant provided samples from healthy sites and diseased sites.

Two studies used DNA-DNA checkerboard (#1, #2) to assess the presence of intracellular or adherent bacteria contrasting with study #3 where fluorescence *in situ* hybridization (FISH) was used. All studies applied probes for the detection of *Porphyromonas gingivalis*, *Aggregatibacter actinomycetemcomitans*, *Tannerella forsythia* and *Treponema denticola*. Study #1 additionally aimed to detect *Prevotella intermedia*, *Prevotella nigrescens*, *Capnocytophaga ochracea*, *Fusobacterium nucleatum subsp. vicentii*, *Campylobacter rectus*, *Veillonella parvula*, *Streptococcus sanguis*, *Streptococcus oralis*, *Streptococcus intermedius* and *Peptostreptococcus micros*. In addition to all these, study #2 also aimed to detect *Actinomyces naeslundii*, *Actinomyces viscosus*, *Actinomyces odontolyticus*, *Actinomyces israelii*, *Actinomyces gerencseriae*, *Capnocytophaga sputigena*, *Capnocytophaga gingivalis*, *Campylobacter showae*, *Eubacterium nodatum*, *Eikenella corrodens*, *Fusobacterium periodonticum*, *Fusobacterium nucleatum subsp. polymorphum*, *Fusobacterium nucleatum subsp. nucleatum*, *Gemella morbillorum*, *Leptotrichia buccalis*, *Neisseria mucosa*, *Propionibacterium acnes*, *Streptococcus anginosus*, *Streptococcus constellatus*, *Streptococcus gordonii*, *Streptococcus mitis* and *Selenomonas noxia*. For global bacteria counts study #3 used DNA probe EUB338. This probe is

universal for eubacteria. The studies #1 and #3 inferred global bacteria counts by the available data but didn't use any specific method. Detailed information regarding the studies characteristics is presented in table 1.

Quality analysis

Methodological quality scores were given according to predetermined criteria (table 2). The highest level of evidence, with low risk of confounding or bias and a moderate probability that the relationship is causal, was attributed to Study #3.

Outcome measurements

Primary outcome - Study #1 found statistically significant differences (Fisher's exact test) regarding the presence of *A. actinomycetemcomitans* (16.7% vs. 0%, $p=0.043$), *T. forsythia* (75% vs. 11.1%, $p<0.001$), *P. intermedia* (54.2% vs. 7.4%, $p<0.001$), *C. ochracea* (37.5% vs. 0%, $p<0.001$) and *C. rectus* (29.2% vs. 0%, $p=0.003$) in epithelial cells from periodontal pockets *versus* healthy sulci. Studies #2 and #3 did not found statistically significant differences regarding specific bacteria.

Secondary outcome - All three studies reported that there were quantitatively more unspecific bacteria in or on the epithelial cells taken from a diseased site when compared with a healthy sulcus, however objective data is only shown in study #3. This study presented a statistically significant higher percentage (chi-square test) of epithelial cells with more than 100 bacteria (>100) in periodontal pockets of subjects with periodontitis compared with epithelial cells from healthy sulci of periodontally healthy subjects (35.8% vs. 0%, $p<0.05$). No statistically significant differences were found for cells harboring 1-20 and 21-100 bacteria. Interestingly, there were also no differences in the percentage of epithelial cells with 1-20, 21-100 or >100 bacteria from periodontal pockets and healthy sulci of periodontitis subjects.

Evidence profile

For quality rating of evidence a several aspects were taken into consideration. The case-control design (observational) of all included studies, the low number of studies, the imprecise or sparse data, the uncertainty about directness and the high risk of bias suggest that results are subject to limitations and therefore must be interpreted with caution (very low quality rating).

DISCUSSION

This systematic review attempted to gather the available evidence on the *in vivo* presence of periodontal pathogens in periodontal tissues and its relationship with periodontal disease.

Summary of main results

All selected studies describe the presence of intracellular and/or adherent bacteria to gingival epithelial cells, *in vivo*, either in periodontal pockets or in healthy sulci. These observations not only corroborate the *in vitro* findings of this bacterial ability⁴⁷⁻⁵², but also sustain that invasion of periodontal tissues occurs even in the presence of a hostile and apparently competent (potential cases with systemic diseases were excluded) immune system. They also substantiate that bacterial invasion, *per se*, is a common event both in health and in disease as suggested by several authors.^{16,20,21,53}

Study #1 found significantly more *A. actinomicetemcomitans*, *T. forsythia*, *P. intermedia*, *C. ochracea* and *C. rectus* intracellularly or adhered to epithelial cells from periodontal pockets when compared to healthy sulci. However, this observation was not supported by the other two case-control studies. In fact, none of these two latter studies found a statistically significant difference in any of the putative periodontal pathogens when comparing health and disease. The lack of consistency in the findings could result from different methodological approaches regarding microbial identification and case and control definitions. Studies #1 and #2 used the

DNA-DNA checkerboard technique. In this technique a positive reaction was recorded only when a signal was greater or equal to (10^4) bacterial cells of the searched specie. This means that only substantial differences in the bacterial load were detected. Study #1 hypothetically potentiated differences by defining cases as having suppuration besides PPD >5 and BOP. A much higher polymorphonuclear neutrophil (PMN) and microbial load in suppurative pockets when compared with healthy sulci⁵⁴ could be a possible explanation for the significant differences found, with such a small sample. Study #2 hypothetically attenuated possible existing differences by comparing periodontal pockets and healthy sulci from the same subjects with periodontitis. It has been recognized that healthy sulcus from subjects with periodontal disease harbor greater number of pathogens when compared with a subject with a healthy *periodontium*.^{55,56} Study #3 used the FISH technique. In this technique a sample was considered positive when at least one epithelial cell in each microscopic field observed presented fluorescent bacteria from the hybridized species-specific probe. Bacterial numbers were estimated by direct counting. Unlike DNA-DNA checkerboard, FISH allows smaller differences to be detected, however as clusters of bacteria are not dispersed the real number of bacteria could be underestimated. Despite all this, all studies seemed to be unanimous in considering that there were quantitatively more bacteria in or on the epithelial cells taken from a diseased site than from a healthy sulcus.

The absence of a significantly higher amount of the 'traditional' periopathogens and the apparent increase of the 'commensal' flora in disease could also be interpreted with reference to the concept of keystone periopathogen described by Hajishengallis.⁵⁷ Accordingly to Hajishengallis, keystone pathogens, specifically *P. gingivalis*, seemed to be able to remodel a symbiotic community into a dysbiotic state by impairing innate immunity in ways that enhanced uncontrolled growth of other species, while being a quantitatively minor constituent of the periodontal microbiota.

Limitations

The studies included in this review are all non-randomized case-control studies. Although having inherent limitations and less level of significance, systematic reviews of observational studies of etiology are especially important. This type of studies are often limited in size and so, only by the simultaneous examination of data from similar studies we can achieve insight into real and spurious associations.⁵⁸

Concerning, however, the scope of this review, the reduced number and the limitations of the included studies preclude a high or even moderate evidence of association between invasion of the *periodontium* by periodontal pathogens and periodontal disease. The current available data is of low to moderate quality and inconsistent mainly due to study design, poor reporting and methodological diversity.

CONCLUSION

There is insufficient evidence to support or exclude the invasion by periodontal pathogens as a key step in the etiopathogenesis of periodontal disease. This review highlights the need of further studies on this topic. Future research should rely on large sample studies, with clear case and control definitions ensuring representativeness of cases and comparability of cases and controls. It's also important to ascertain exposure to invasive periodontal pathogens with maximum detail (e.g. strains, intracellular/intercellular, synergisms) using a blinded operator.

METHODS

This systematic review was conducted in accordance with the guidelines of Cochrane Collaboration⁵⁹ and Transparent Reporting of Systematic Reviews and Meta-Analyses [PRISMA statement].⁶⁰ The focused question was given in the following: What is the relationship between the quality or quantity of putative periodontal pathogens in periodontal tissues and periodontal disease?

Eligibility criteria

All studies that reported evaluation of periodontal pathogens invasion on human tissues were considered eligible. To be as inclusive as possible, no restrictions were applied with regard to the year of publication or to language. Exclusion decision of non-English papers was delayed until the next step. Reviews and case reports were excluded.

Search Strategy

Four Internet sources of evidence were used in search of appropriate papers that fulfill the purpose of the study: the National Library of Medicine, Washington D.C. (MEDLINE-PubMed), the Cochrane Central Register of Controlled Trials (CENTRAL), ISI Web of Knowledge (Thomson Reuters) and SCOPUS (Elsevier). The databases were searched up to 22nd of January, 2014. A structured search strategy was developed to include any published paper that investigated the invasion of periodontal pathogens in patients with periodontal disease. The following strategy was used in the search: (((periodontitis OR “periodontal disease” OR “periodontal pocket”) AND (identification OR detection OR localization) AND bacteria*) AND invas* OR intracellular OR tissue* OR “epithelial cells”). Key words were combined with Boolean operators and the asterisk symbol (*) was used as truncation. The search strategy was customized according to the database being used.

Screening and selection

Firstly, two independent reviewers (LM and MP) screened titles and abstracts for eligible papers. Secondly, the full texts of those papers were obtained and screened in relation to the study purpose, protocol and reported data. An additional hand search across the reference list of the selected studies was made as an attempt to find additional papers that could meet the eligibility criteria.

Data extraction and analysis

Data from the papers that met the selection criteria were processed for analysis. Information regarding year of publication, study design, population, inclusion and exclusion criteria, case and control definitions, characteristics of participants, type of intervention, ascertainment of exposure, cell sample, microorganisms found and its location and statistical analysis were collected by LM e MP using data extraction sheets.

Heterogeneity of trials

The heterogeneity across the trials was detailed according to the following factors: population characteristics, study design, case and control definition, used methodology and type pathogens found.

Quality assessment

The methodological quality of the selected studies was independently scored by two reviewers (LM and MP). Any disagreement was resolved after additional discussion. As all eligible papers were non-randomized studies (case-control studies) methodological quality was assessed combining several proposed criteria of the STROBE statement⁶¹, the Newcastle – Ottawa quality assessment scale⁶², the Downs and Black checklist for non-randomized studies⁶³ and SIGN50 guidelines⁶⁴ as suggested by the Quality Assessment Tools Project Report elaborated by the Canadian Agency for Drugs and Technologies in Health.⁶⁵ In short, a study was classified as high quality with a very low risk of confounding or bias and a high probability that the relationship is causal (++) when all of the following criteria were adequately addressed: appropriate and clearly focused question, defined exclusion criteria, case selection, control selection, representativeness of cases, comparability of cases and controls, ascertainment of exposure, blindness, funding and statistical analysis. When some of the criteria have not been fulfilled or

not adequately described but this fact is thought unlikely to alter the conclusions the study was classified as well conducted study with low risk of confounding or bias and a moderate probability that the relationship is causal (+). When some of the criteria have not been fulfilled or not adequately described and this fact is thought likely to alter the conclusions (-) the study was classified as a study with a high risk of confounding or bias and a significant risk that the relationship is not causal (-).

Outcome variables

Primary outcome measures of interest were intracellular or adherent (to cell /periodontal tissues) mean value or percentage of known pathogens. Secondary outcomes of interest were global counts of intracellular or adherent (to cell /periodontal tissues) unspecific bacteria.

Evidence profile

The Grading of Recommendations Assessment, Development and Evaluation (GRADE) system⁶⁶ as proposed by the GRADE working group was used for grading the collective evidence emerging from this review. Two reviewers (LM and MP) rated the quality of the evidence for outcomes across studies. Any disagreement was resolved after additional discussion.

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FIGURES

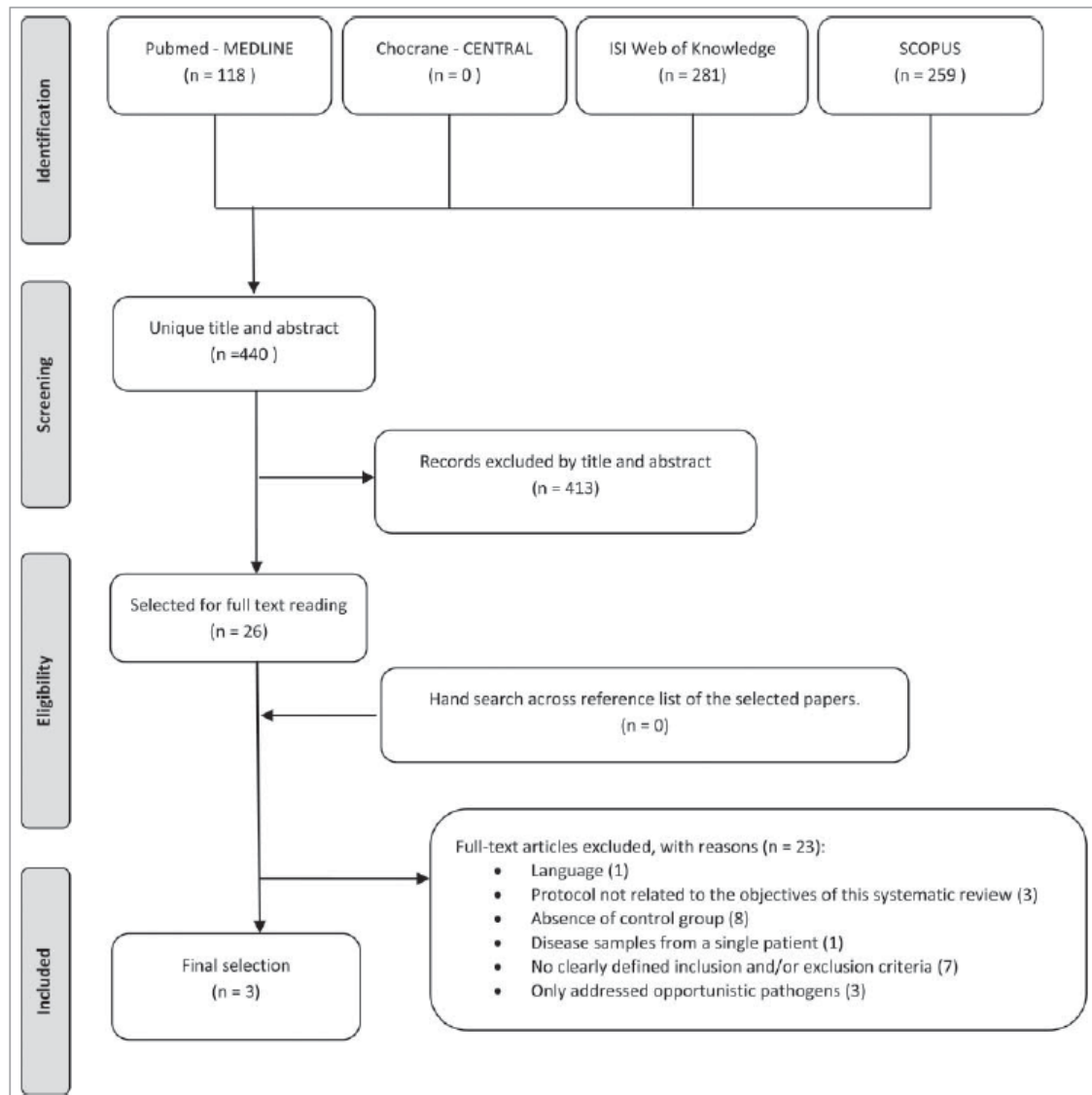


Fig. 1. Flowchart of literature search and study selection

TABLES

Table 1. Characteristics of included studies

#	Study design	A. Subject characteristics			
		n	Population	Male/Female%(age±SD)	Smokers (%)
1	Dibart et al. (1998) Case-control	Cases	24 Patients from Center for Clinical Research in Periodontal Disease	Unknown	Unknown
		Control	27 Periodontal healthy dental students		
2	Colomb o et al. (2006) Case-control	Cases	120 Patients with moderate to severe CP from Federal University of Rio de Janeiro	43/57% (46.3±1.4)	Non-smokers (51%) Past smokers (27%) Smokers (22%)
		Controls	92		
3	Colomb o et al. (2007) Case-Control	Cases	175 Patients with CP from Federal University of Rio de Janeiro	57/43% (44.2 ±1.4) 21/79 % (29.9 ± 2.3)	Unknown
		Controls	68 Periodontal healthy subjects		

PPD = Probing pocket depth (mm); CAL = Clinical attachment loss (mm); PI= Plaque index (%); BOP = Bleeding on probing

Table 1. Characteristics of included studies

B. Intervention characteristics					
#	Sample	Sample site characteristics	Intervention	Microorganisms	Microbiological differences <i>Bacteria (disease vs. health, p)</i>
1	Epithelial cells from periodontal pockets	Cases PPD= 6.5	DNA-DNA checkerboard	<i>P. gingivalis</i> , <i>A. actinomycetemcomitans</i> , <i>T. forsythia</i> , <i>T. denticola</i> , <i>P. intermedia</i> , <i>P. nigrescens</i> , <i>C. ochracea</i> , <i>F. nucleatum</i> subsp. <i>vicentii</i> , <i>C. retus</i> , <i>V. parvula</i> , <i>S. sanguis</i> , <i>S. oralis</i> , <i>S. intermedius</i> and <i>P. micros</i>	Aa. (16.7% vs. 0%, p=0.043) Tf. (75% vs. 11,1%, p<0.001) Pi. (54.2% vs. 7.4%, p<0.001) Co. (37.5% vs. 0%, p<0.001) Cr. (29.2% vs. 0%, p=0.003)
		Controls PPD = 2.5			
2	Epithelial cells from periodontal pockets	Cases PPD = 5.8 ± 0.3 CAL = 6.8 ± 0.3	DNA-DNA checkerboard	<i>P. gingivalis</i> , <i>A. actinomycetemcomitans</i> , <i>T. forsythia</i> , <i>T. denticola</i> , <i>P. intermedia</i> , <i>P. nigrescens</i> , <i>C. ochracea</i> , <i>F. nucleatum</i> subsp. <i>vicentii</i> , <i>C. retus</i> , <i>V. parvula</i> , <i>S. sanguis</i> , <i>S. oralis</i> , <i>S. intermedius</i> , <i>P. micros</i> , <i>A. naeslundii</i> , <i>A. viscosus</i> , <i>A. odontolyticus</i> , <i>A. israelii</i> , <i>A. gerencseriae</i> , <i>C. sputigena</i> , <i>C. gingivalis</i> , <i>C. showae</i> , <i>E. nodatum</i> , <i>E. corrodens</i> , <i>F. periodonticum</i> , <i>F. nucleatum</i> subsp. <i>polymorphum</i> , <i>F. nucleatum</i> subsp. <i>nucleatum</i> , <i>G. morbillorum</i> , <i>L. buccalis</i> , <i>N. mucosa</i> , <i>P. acnes</i> , <i>S. anginosus</i> , <i>S. constellatus</i> , <i>S. gordonii</i> , <i>S. mitis</i> and <i>S. noxia</i> ,	No statistically significant differences regarding specific bacteria
		Controls PPD = 1.8 ± 0.1 CAL = 2.3 ± 0.1			
3	Epithelial cells from periodontal pockets, gingival crevice and buccal mucosa	Cases PPD = 5.8 ± 0.3 CAL = 6.8 ± 0.3 PI = 86 ± 11 BOP = 70 ± 18	Fluorescence in situ hybridization laser-scanning confocal microscopy	<i>P. gingivalis</i> , <i>A. actinomycetemcomitans</i> , <i>T. forsythia</i> and <i>T. denticola</i> .	No statistically significant differences regarding specific bacteria GBC (35.8% vs. 0%, p<0.05).
		Controls PPD = 1.8 ± 0.1 CAL = 2.3 ± 0.1 PI = 27 ± 15 no BOP			

PPD = Probing pocket depth (mm); CAL = Clinical attachment loss (mm); PI= Plaque index (%); BOP = Bleeding on probing; Aa.=A. actinomycetemcomitans; Tf.=T. Forsythia; Pi.=P. intermedia; Co.= C. ochracea; Cr.= C. retus; GBC= global bacterial counts in or on epithelial cells (>100).

Table 2. Quality assessment of included studies

#		<i>Appropriate and clearly focused question</i>	<i>Defined Exclusion Criteria</i>	<i>Case Selection</i>	<i>Control Selection</i>	<i>Representativeness of cases</i>	<i>Comparability of cases and controls</i>	<i>Ascertainment of Exposure</i>	<i>Blindness</i>	<i>Statistical Analysis</i>	<i>Funding</i>	<i>Overall assessment</i>
1	Dibart et al. (1998)	2	2	1	1	1	0	1	0	1	2	-
2	Colombo et al. (2006)	2	2	2	1	1	2	1	0	1	2	-
3	Colombo et al. (2007)	2	2	2	1	1	2	2	0	1	2	+

(0 = not addressed, 1 = poorly addressed, 2 = adequately addressed)

(++ All or most of the criteria have been fulfilled. Where they have not been fulfilled the conclusions of the study or review are thought very unlikely to alter; + some of the criteria have been fulfilled. Those criteria that have not been fulfilled or not adequately described are thought unlikely to alter the conclusions; - some of the criteria have been fulfilled. Those criteria that have not been fulfilled or not adequately described are thought likely to alter the conclusions.)

